

Analysing the genomic complexity at the single cell level using Oxford Nanopore Technology

BY: VARSHA BALRAJ (z5113220)

Supervisors: A/Prof. Fabio Luciani ,
Dr Preston Leung

Thesis B Presentation

Outline

- Project Recap
- Thesis B
 - Changes in method
- Current Progress
 - Results
- Timeline

Project Recap

From Thesis A

Problem

- There is no research to study isoforms occurring in T cell data.
- Previous technology made study of isoforms almost impossible due to the estimation involved

Aim

To study single cell data with Oxford Nanopore sequencing to understand the different isoforms of CD45 in T cells

Hypothesis

Analysis of single cells data with long reads can be used to identify multiple isoforms present in the CD45 gene.

Thesis B Aims

- Align sequences to reference
- Understanding the alignment
- Calculate reads to each isoform
- Identify combination of exons that make up each known isoform

Previous method

Option 1

Download Isoform Data for CD45 from Ensemble

Combine exons of each isoform into a file

Isoform Dataset

Clinical Samples

Single Cell Sequencing + Oxford Nanopore Sequencing

Reference Data

Option 2

CANU Assembly

De Novo Method
Hierarchical assembly pipeline
<https://github.com/marbl/canu>

Sequence Alignment

MiniMap2
<https://www.ncbi.nlm.nih.gov/pubmed/29750242>
Bowtie2
<http://bowtie-bio.sourceforge.net/bowtie2/manual.shtml>

Quantify Data

Count number of reads to each isoform
Identify combination of exons that make up each isoform

Visualisation of Data

Analysis of Data

Thesis B

Problems with previous method

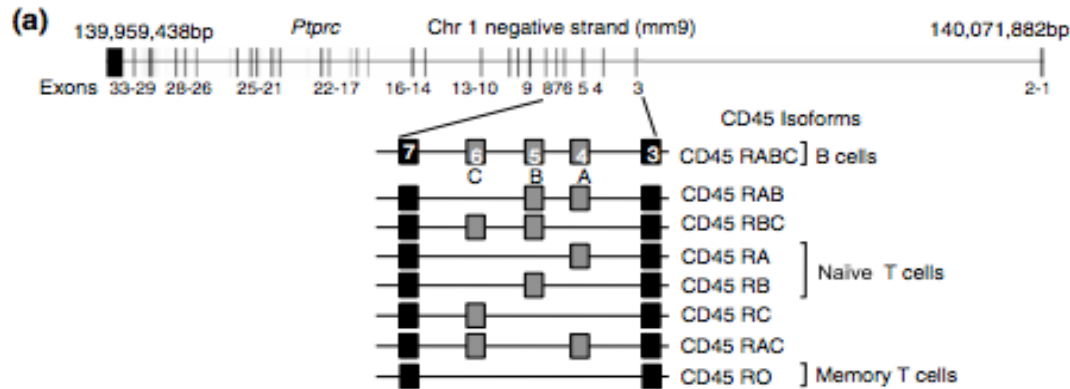
- When downloaded the isoform data, most sequences were not complete
- Not able to form a complete isoform reference dataset

New Approach

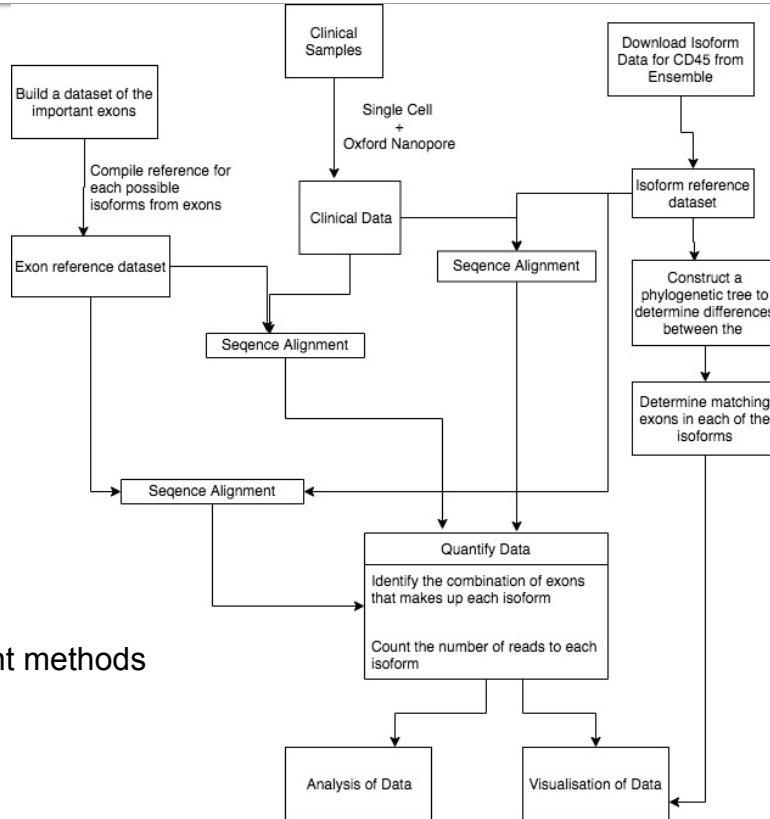
- Focus on the exons themselves in detail
- Notice the differences between the isoforms from the exons
- Build a reference data set from exons only

Exons in more detail

- Based on the paper (*Vicky Cho, Genome Biology 2014, 15:R26*)



Refined Method

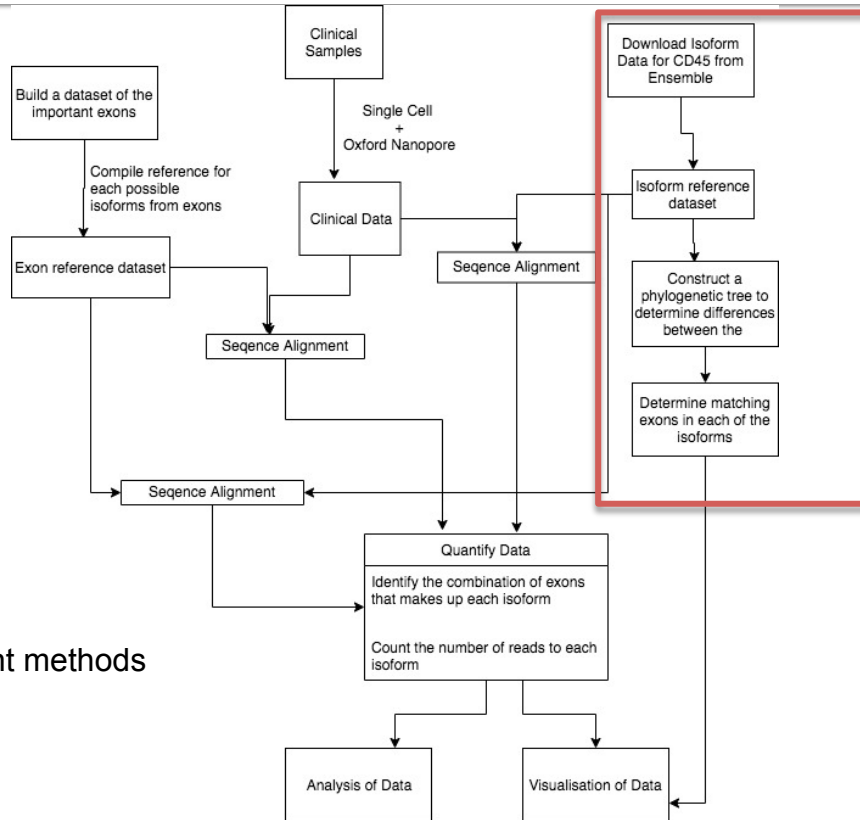


Note: Sequence alignment methods
 -Minimap2
 -Bowtie2

Current Progress

Results

Refined Method

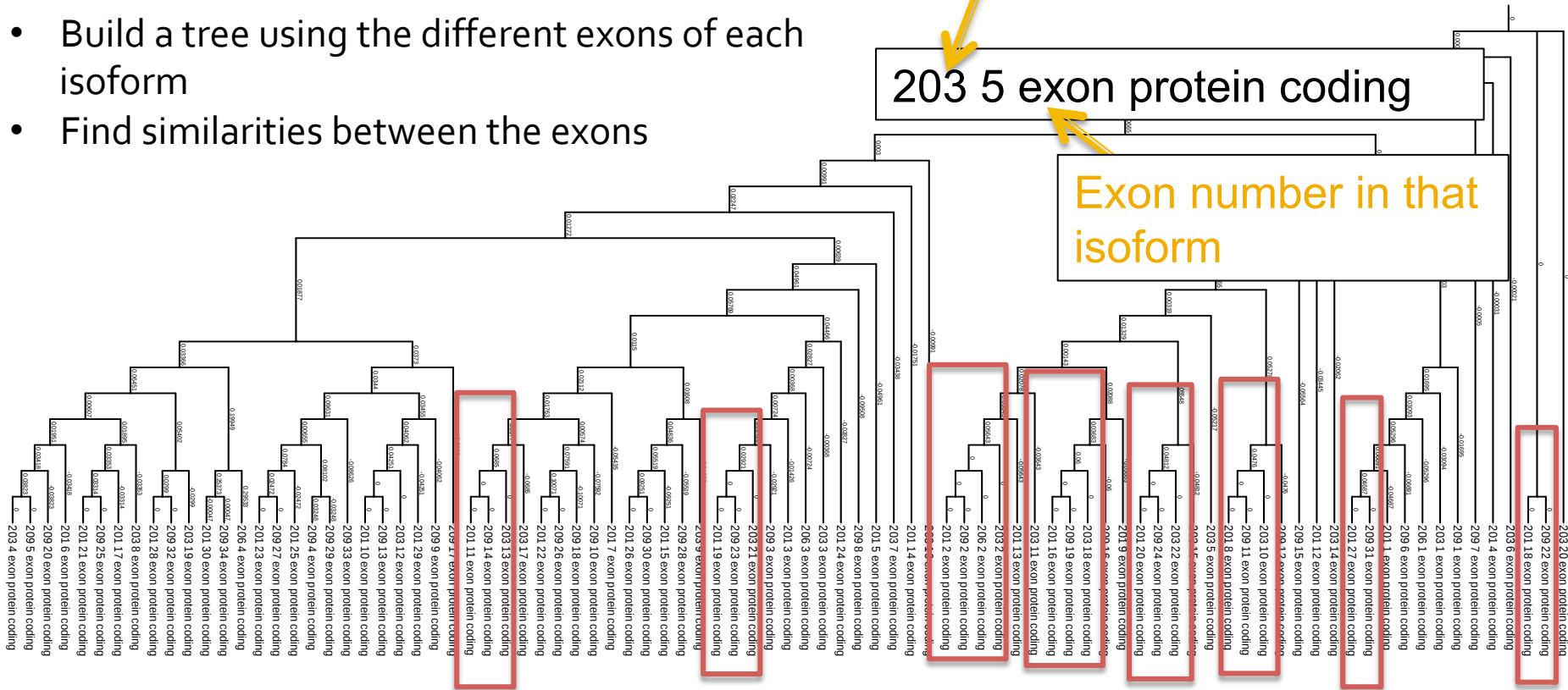


Note: Sequence alignment methods
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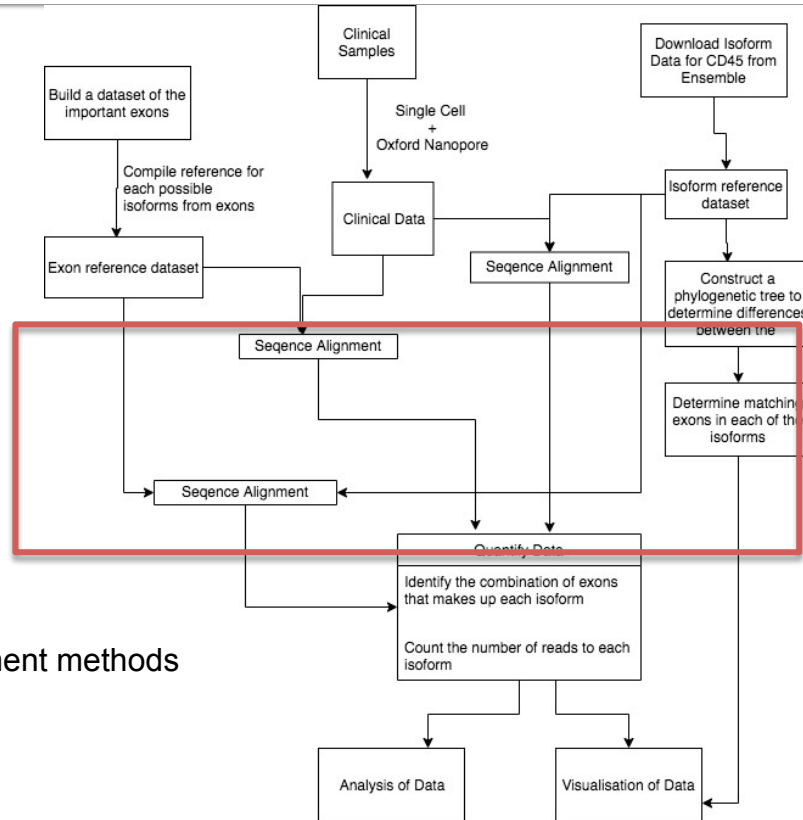
Phylogenetic Tree

Isoform number

- Build a tree using the different exons of each isoform
- Find similarities between the exons



Refined Method



Note: Sequence alignment methods
-Minimap2
-Bowtie2

Sequence Alignment

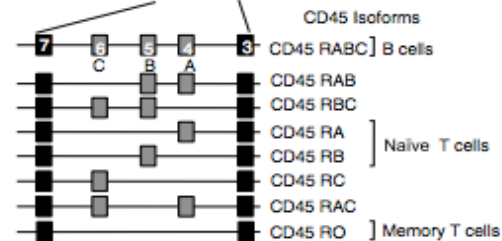
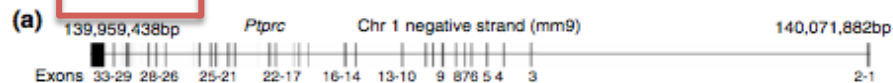
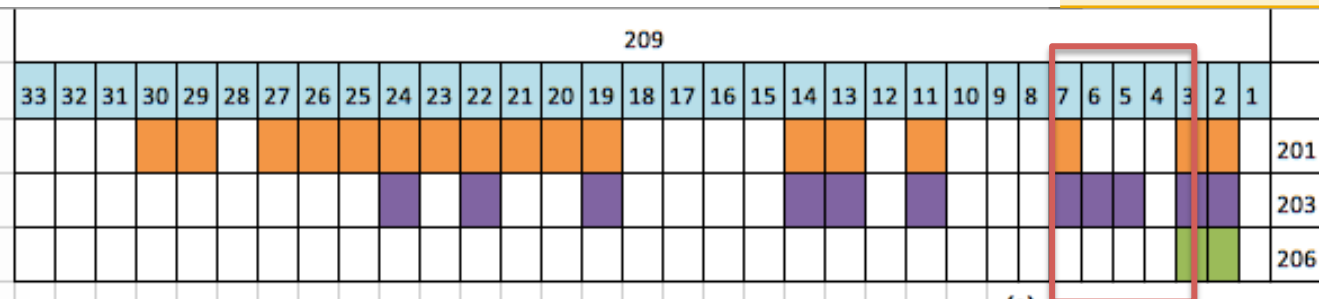
- Align exons to the four isoforms using minimap2

Isoforms	Exons					
	2	3	4	5	6	7
201	Y	Y	N	N	N	Y
203	Y	Y	N	Y	Y	Y
206	Y	Y	N	N	N	N
209	Y	Y	Y	Y	Y	Y

Mapping exons in each Isoform

- Construct a map for isoforms and corresponding exons
- Narrowing into genomic exons 4, 5, 6

Isoform	201	203	206	209
Total no. of Exons	30	22	4	33



Candidate Isoforms

209

CDRABC

206

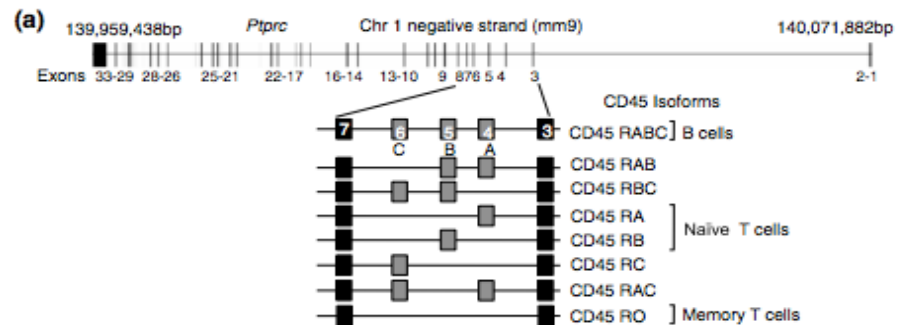
Very little-
almost no
alignments

203

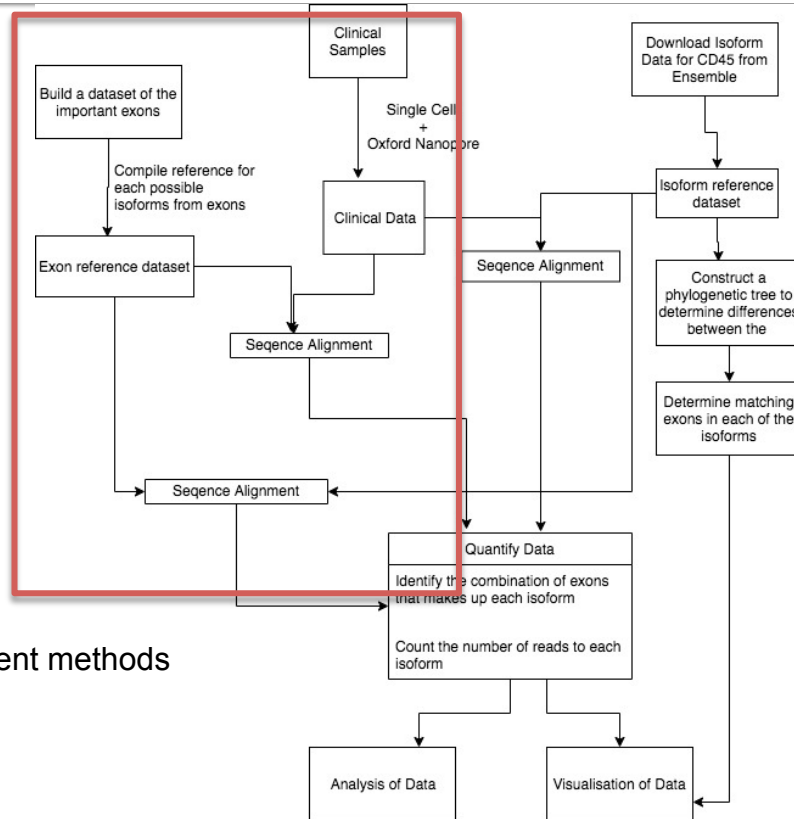
CDRBC

201

CDRO



Refined Method



Note: Sequence alignment methods

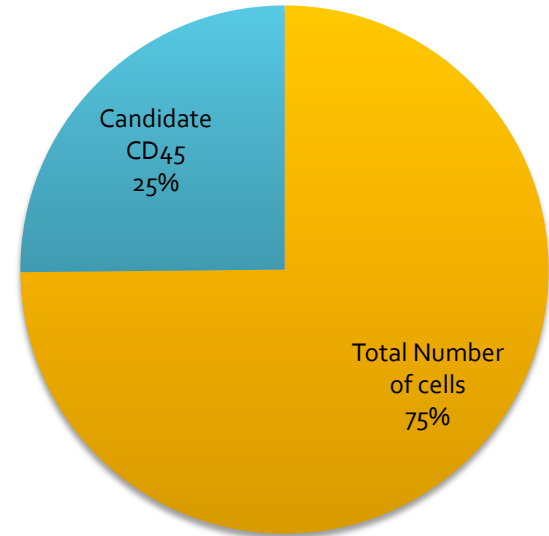
-Minimap2

-Bowtie2

Experimental measurement of CD45

- Results of the alignment of N=476 for which experimental measurements of CD45 protein expression was available
- N=160 (25%) had contigs that aligned to ANY reference isoform .
- These candidate cells aligned to either 209 (all the exons) or exon 2& 3 &7.

Total Number of cells	Candidate CD45
476	160

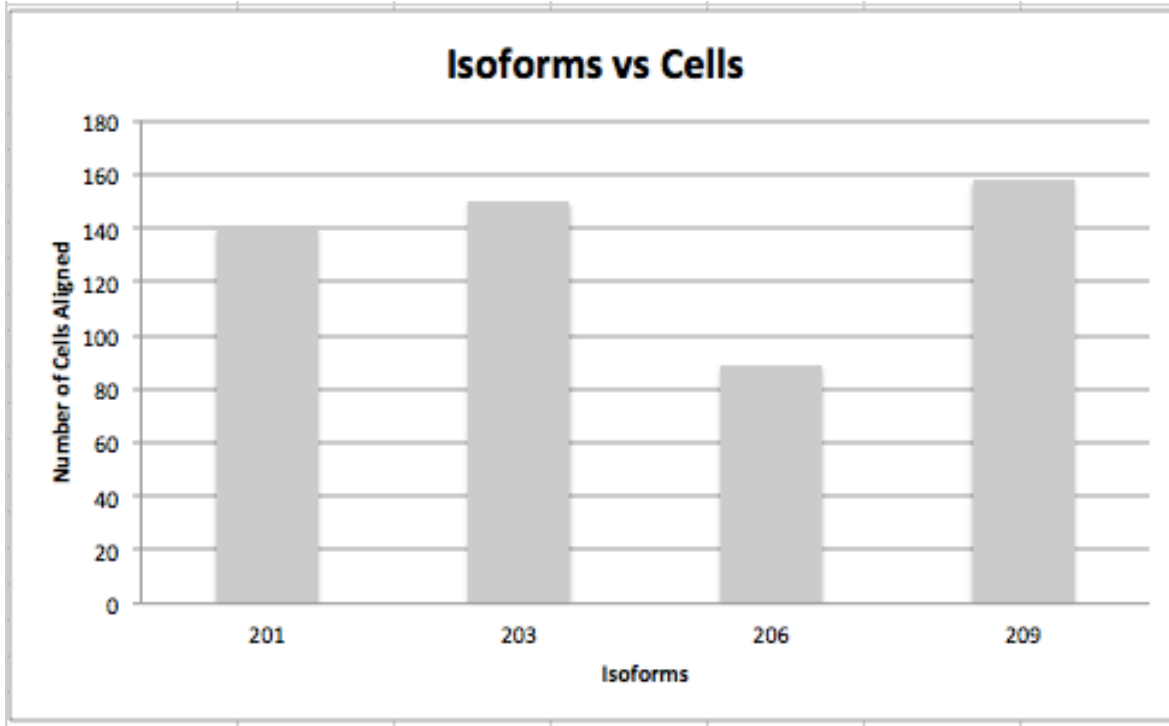


RESULTS shown are from alignment with references (using minimap2 aligner)

Applying to Cell Data

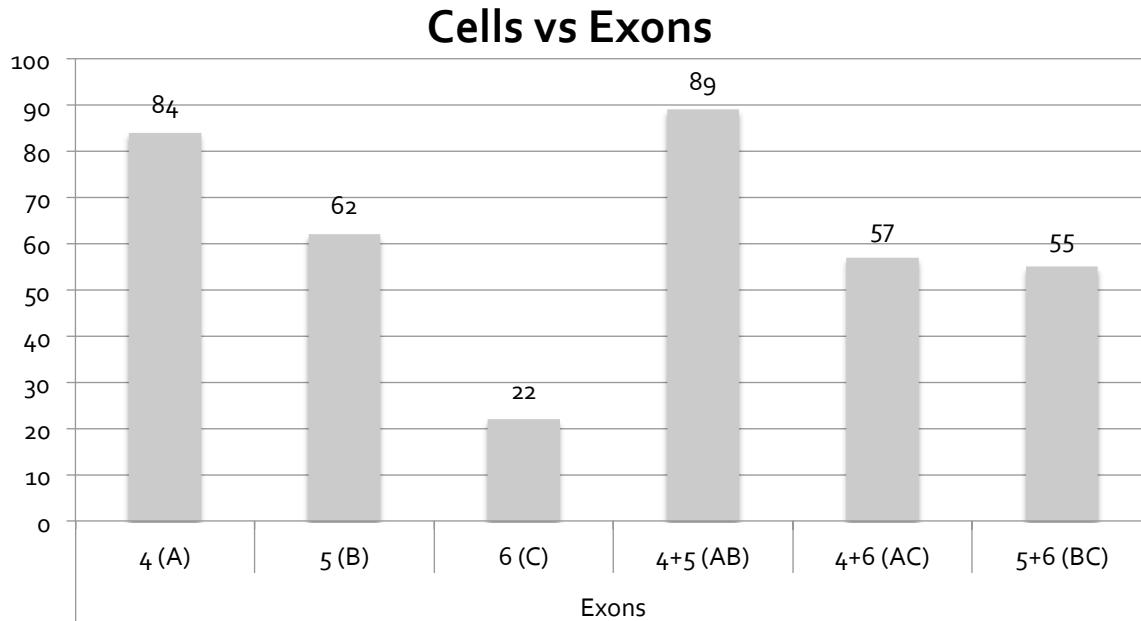
- Total number of candidate cells aligned = 160

Isoforms	201	203	206	209
No. Cells Aligned	140	150	89	158



Mapping of contigs within Exon & Cell Alignment

	Exons					
	4 (A)	5 (B)	6 (C)	4+5 (AB)	4+6 (AC)	5+6 (BC)
Num Cells Aligned	84	62	22	89	57	55



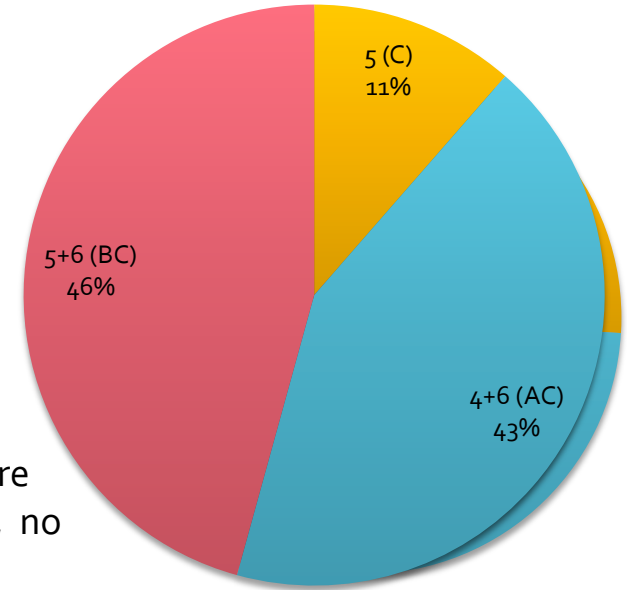
Individual Exons

- Total number of Candidate CD45 cells
N = 160

	5 (C) 5 (B)	4+6 (AC) 4+5 (AB)	5+6 (BC) 5+6 (BC)
Num Cells Aligned	84	89	57
Num Cells Aligned	62	89	55
Uniquely	27	58	16

- Unique – exon that was present but no other exons of interest were
- Unique Combination – only exons the present 2 exons aligned, no traces of 3rd

Unique Distribution of Exon 5
Unique Distribution of Exon 6

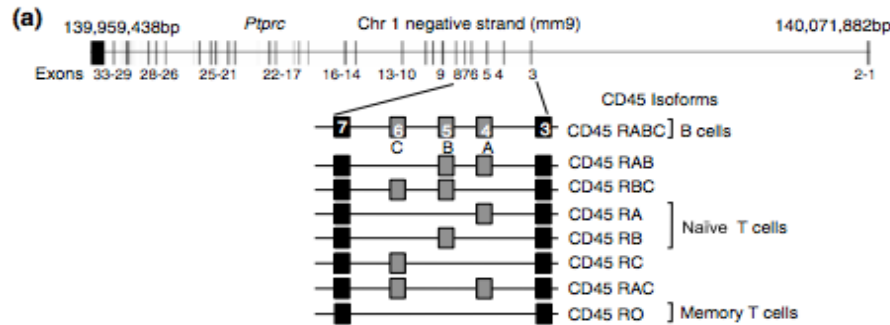
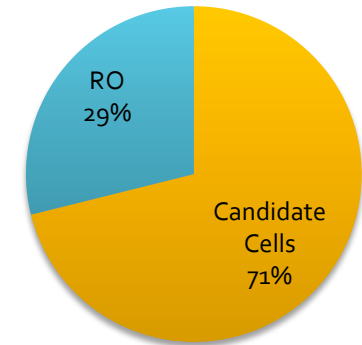


Finding RO

- Known CD45 cells that did not align to exons 4, 5, 6

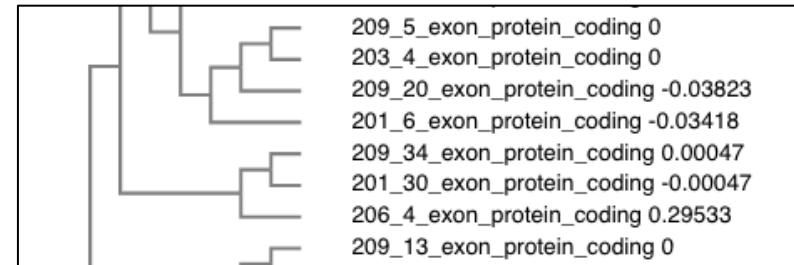
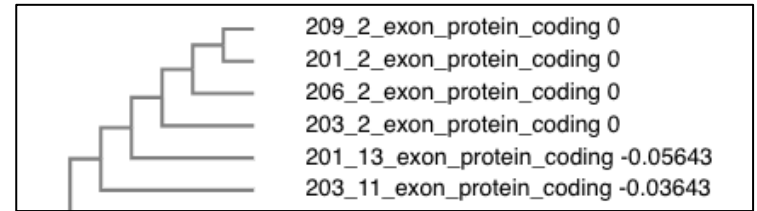
Candidate Cells	RO
160	65

Candidate RO Cells

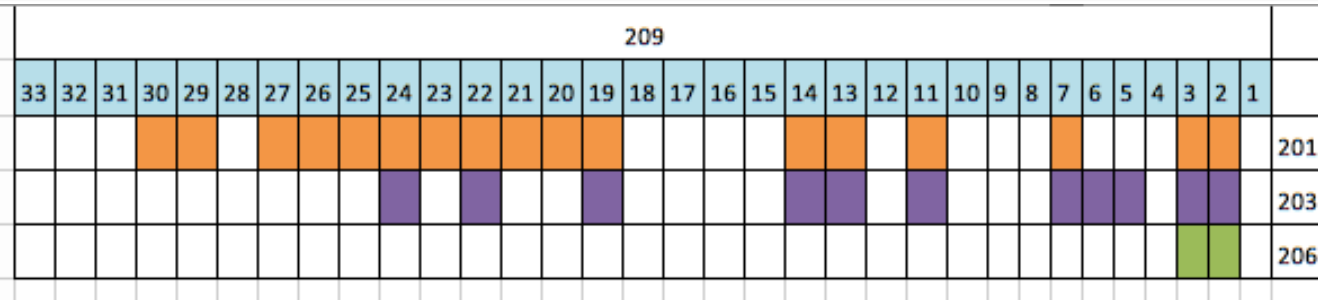


Other interesting findings

- Some exons in the same isoforms are very similar
- Potential repeating segments?



Other interesting findings



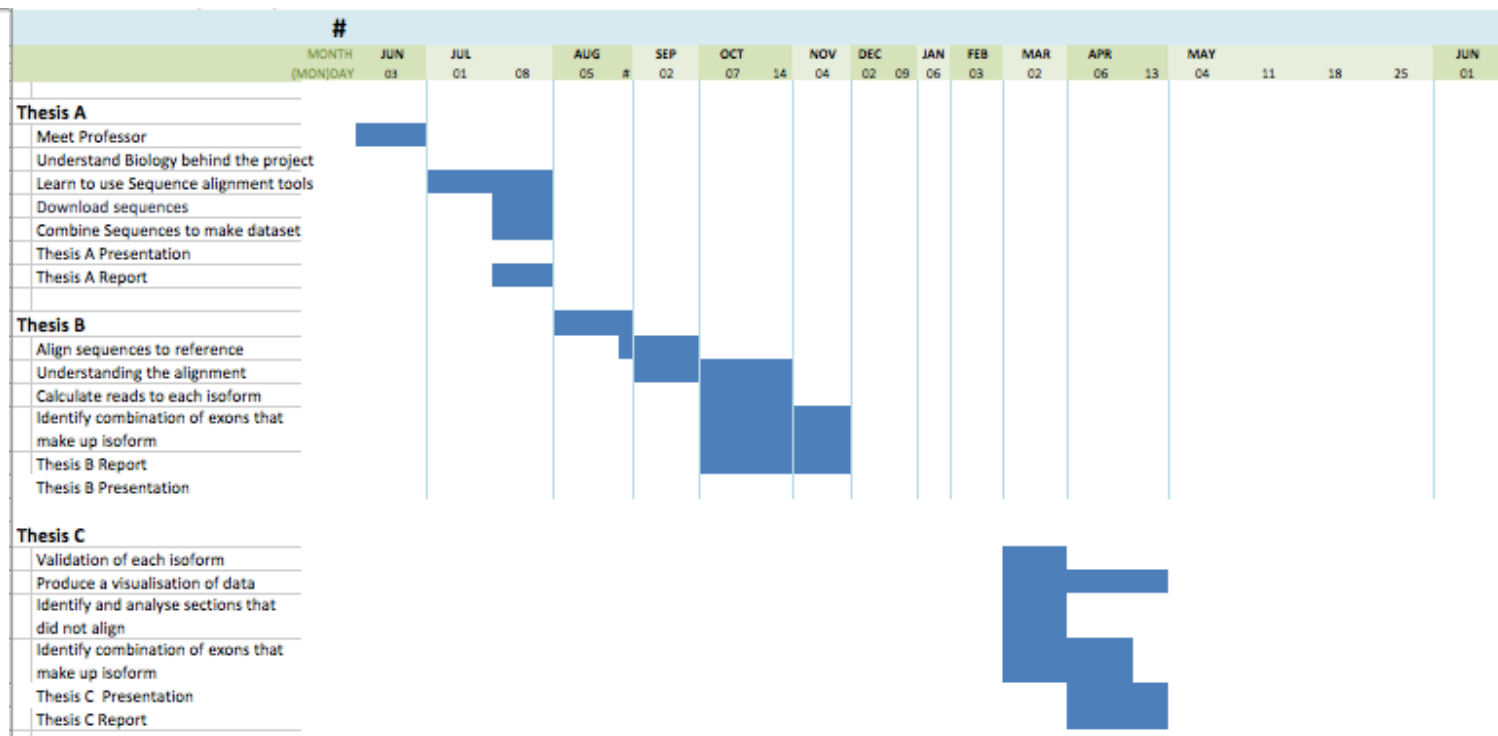
- What lies beyond the exons of interest (exon 4, 5, 6)
- 33 known exons – functions of the other exons and if they contribute to isoforms as well?

Limitations and future directions

- Isoform alignments produced more alignments, overlaps of other 30 exons also playing a part as variables
- Used only protein coding sequences
- Results will need to be validated by comparing protein expression of RA & RO

Project Timeline

Project Timeline



Thesis C

- Validation of each isoform
- Produce a visualisation of data
- Investigate other 30 exons
- Investigate similarities between exons

References

In-text: (Longdom.org, 2019)

Your Bibliography: Longdom.org. (2019). [online] Available at: <https://www.longdom.org/open-access/generations-of-sequencing-technologies-from-first-to-next-generation-0974-8369-1000395.pdf>

In-text: (Illumina.com, 2019)

Your Bibliography: Illumina.com. (2019). [online] Available at: https://www.illumina.com/documents/products/illumina_sequencing_introduction.pdf [Accessed 20 Jul. 2019].

In-text: (Biorxiv.org, 2019)

Your Bibliography: Biorxiv.org. (2019). [online] Available at: <https://www.biorxiv.org/content/biorxiv/early/2019/01/28/530824.full.pdf> [Accessed 20 Jul. 2019].

In-text: (Anon, 2019)

Your Bibliography: Anon, (2019). [online] Available at: <https://www.sciencedirect.com/topics/neuroscience/nanopore-sequencing> <https://bitesizebio.com/36592/introduction-nanopore-sequencing/> [Accessed 20 Jul. 2019].

Your Bibliography: Anon, (2019). [online] Available at: <https://www.sciencedirect.com/topics/neuroscience/nanopore-sequencing> <https://bitesizebio.com/36592/introduction-nanopore-sequencing/> [Accessed 20 Jul. 2019].

DNA SEQUENCING FACT SHEET

In-text: (Genome.gov, 2019)

Your Bibliography: Genome.gov. (2019). *DNA Sequencing Fact Sheet*. [online] Available at: <https://www.genome.gov/about-genomics/fact-sheets/DNA-Sequencing-Fact-Sheet>

SINGLE CELL DNA SEQUENCING | NOVOGENE

In-text: (Novogene, 2019)

Your Bibliography: Novogene. (2019). *Single Cell DNA Sequencing | Novogene*. [online] Available at: <https://en.novogene.com/next-generation-sequencing-services/single-cell-sequencing/single-cell-dna-sequencing/>

SINGLE-CELL AND LOW-INPUT RNA-SEQ | SINGLE-CELL SEQUENCING BENEFITS

In-text: (Sapac.illumina.com, 2019)

Your Bibliography: Sapac.illumina.com. (2019). *Single-Cell and Low-Input RNA-Seq | Single-cell sequencing benefits*. [online] Available at: <https://sapac.illumina.com/techniques/sequencing/rna-sequencing/ultra-low-input-single-cell-rna-seq.html> [Accessed 20 Jul. 2019].